

EXPERIMENTAL

Isolation The air-dried capsules (without seeds, 640 g) of *M. japonicus* Muell Arg were extracted with MeOH at room temp for 4 days. The MeOH filtrate was concd to give ppts, which were crystallized from MeOH to afford yellow needles (compound A). The filtrate separated from A was evaporated to dryness, dissolved in H₂O and extracted with hexane and then with EtOAc to give a hexane extract (9.7 g) and an EtOAc extract (24.3 g). The latter was purified on a Sephadex LH-20 column (CHCl₃-MeOH, 1:1) followed by repeated CC on Si gel (hexane-CHCl₃ and then hexane-EtOAc) to afford additional A and compound B.

Compound A Yellow needles (2.4 g) from MeOH, mp 188–189° (uncorr). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(ϵ) 293 (23 200), IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹, 3310 (OH), 3220, 1615, 1595, 1558, 1434, 1390, 1365, 1260, 1208, 1172, 1128, MS m/z 444 [M]⁺, 389 [M - 55]⁺, 249, 235, 209, 195, 181, NMR Tables 1 and 2 (Found C, 64.35, H, 6.44 C₂₄H₂₈O₈ requires C, 64.85, H, 6.35%) **Pentamethyl ether of A** A MeOH soln of A (14.1 mg) was treated with CH₃N₂-Et₂O. The resulting products purified by CC on Si gel gave a colourless oil (4.2 mg), MS m/z 514 [M]⁺, ¹H NMR δ (CDCl₃) 1.68, 1.75 (3H, br s each, Me), 2.15 (3H, s, Me), 2.51 (6H, s, Ac), 3.31 (2H, d, J = 6.2 Hz), 3.46, 3.59 (3H, s each, OMe), 3.50, 3.70 (6H, s each, OMe), 4.00 (2H, s, CH₂ between rings), 5.16 (1H, t, J = 6.2 Hz) **Penta-acetate of A** A (20 mg) was acetylated with Ac₂O (1 ml) and pyridine (1 ml) at room temp for 10 min. After usual treatment the resulting products were purified by CC on Si gel. A colourless oil (15 mg) was obtained, MS m/z 654 [M]⁺, ¹H NMR Table 1.

Compound B Yellow needles (88 mg) from MeOH, mp 197–199° (uncorr). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(ϵ) 232 (22 600), 289 (18 600), 323 (16 400), IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹ 3320 (OH), 3230, 1603, 1415, 1362, 1292, 1170, 1128, MS m/z 460 [M]⁺, 417 [M - 43]⁺, 389 [M - 71]⁺, 209, 195, 181, NMR Tables 1 and 2 (Found C,

62.34, H, 6.23 C₂₄H₂₈O₉ requires C, 62.60, H, 6.13%) **Hexa-acetate of B** Using acetylation as described for A, a colourless oil was obtained, MS m/z 712 [M]⁺, ¹H NMR Table 1.

Reductive alkaline cleavage of A A (120 mg) dissolved in 5% NaOH (60 ml) was mixed with Zn powder (0.6 g) and warmed for 5 min at 100°. The filtrate of the reaction mixture was acidified with 10% H₂SO₄ and extracted with Et₂O. After evaporation of Et₂O the residue was purified through a Si gel column (hexane-EtOAc, 13:5) to afford yellow needles (32.5 mg), mp 197–200° (uncorr) MS m/z 196 [M]⁺, 181, ¹H NMR (CD₃OD) δ 1.91 (3H, s, Me), 2.63 (3H, s, Ac), 3.81 (3H, s, OMe), 6.00 (1H, s). It was identified by comparison with an authentic sample of 2, 6-dihydroxy-3-methyl-4-methoxyacetophenone [6] by mmp and comparison of spectral data.

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CANDIDOL, A FLAVONOL FROM *TEPHROSIA CANDIDA*

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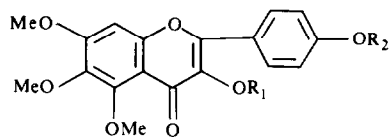
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Abstract—The seeds of *Tephrosia candida* have yielded a new flavonol, characterized here as 3,4'-dihydroxy-5,6,7-trimethoxyflavone.

Earlier investigations [1, 2, Chibber, S. S. and Dutt, S. K., unpublished] of the seeds of *Tephrosia candida* have revealed the presence of three new flavonoids. We report here the isolation and characterization of a

flavonol, candidol, from the ethyl acetate extract of the seeds. It analysed for C₁₈H₁₆O₇ and produced a yellow fluorescence in UV light. It responded to Shinoda's test for flavonoids giving a magenta colour.



1 $R_1 = R_2 = H$

2 $R_1 = R_2 = Me$

and also gave a brownish-green colour with alcoholic ferric chloride indicating a chelated hydroxyl group. The UV spectrum showed λ_{max} in methanol at 270 and 355 nm suggestive of a flavonol. The presence of a chelated hydroxyl was supported by a bathochromic shift of 67 nm with aluminium chloride–hydrochloric acid in UV and absorptions at 3430 (ν_{OH}) and 1640 (ν_{CO}) cm^{-1} in the IR spectrum. Absence of a shift with sodium acetate indicated the absence of a free hydroxyl at the 7-position. However, a bathochromic shift of 35 nm with sodium methoxide suggested a 4'-hydroxyflavonol skeleton [3]. This was substantiated by two doublets ($J = 10$ Hz) at δ 7.00 and 8.05 in the 1H NMR spectrum, characteristic of an A_2B_2 pattern, due to the B-ring protons. The 1H NMR spectrum further showed the presence of three methoxyl groups which were indicated by two singlets at δ 3.70 and 4.00, integrating for three and six protons, respectively. A sharp singlet at δ 6.64 integrating for one proton was indicative of the fact that only one position in the A-ring was unsubstituted. Absence of any low field deshielded proton suggested that the 5-position was substituted. Methylation of candidol yielded a compound which was identified as 3,5,6,7,4'-pentamethoxyflavone (2) [4]. This established the oxygenation pattern in candidol and showed the presence of two hydroxyl groups in addition to the three methoxys. The second hydroxyl was assigned to the 3-position on the basis of a negative Gibbs test. Hence all three methoxys must be present in ring A and this was confirmed by oxidation of the dimethyl ether (2), when 2-hydroxy-4,5,6, ω -tetramethoxyacetophenone and anisic acid were obtained. The former was identified by direct comparison with an authentic sample prepared by the method of Row and Seshadri [5]. A retro-Diels–Alder fragmentation pattern was not observed in the mass spectrum and is in agreement with the observation that such a process is of minor importance in the highly substituted flavones [6, 7]. The structure 3,4'-dihydroxy-5,6,7-trimethoxyflavone (1) for candidol, assigned on the basis of spectral and chemical data, was finally confirmed by comparing it with a synthetic sample [8] (mmp and co-TLC).

It is interesting to note that 6-hydroxykaempferol

4'-methyl ether 3,7-dihydroxyflavone, a flavonol glycoside with the same substitution pattern, has been isolated from the aerial parts of *T. candida* [4].

EXPERIMENTAL

Isolation Air-dried, coarsely powdered seeds (1 kg) of *T. candida* were extracted with EtOAc after exhaustive treatment with petrol to remove petrol-soluble oils. This extract was concd under red pres and subjected to CC over Si gel. On elution with C_6H_6 –EtOAc (8/2) it gave a pale yellow, crystalline compound (80 mg, mp 253–254°), candidol (1). TLC R_f 0.16 (C_6H_6 –EtOAc, 4/1), 0.60 (C_6H_6 –EtOAc, 1/1). UV λ_{max}^{MeOH} nm 270, 340, 355, +NaOAc 270, 340 +NaOMe 270, 390, +AlCl₃ 235, 275, 345, 422, +AlCl₃–HCl 235, 285, 360, 422. IR ν_{max}^{KBr} cm^{-1} 3430, 1640, 1590, 1460, 995 and 800. 1H NMR (90 MHz, CDCl₃) δ 3.70 (3H, s, OMe), 4.00 (6H, s, 2 \times OMe), 6.64 (1H, s, Ar–H-8), 7.00 (2H, d, $J = 10$ Hz, Ar–H-3', H-5'), 8.05 (2H, d, $J = 10$ Hz, Ar–H-2', H-6'). MS m/z (rel int) 343 [$M - 1$]⁺ (67), 326 (24), 301 (19), 286 (16), 258 (10), 181 (61), 167 (21), 153 (100), 134 (97) and 121 (97). Methylation (Me_2SO , K_2CO_3 , Me_2CO) gave the dimethyl ether as brown needles, mp 151–153°, characterized as 3,5,6,7,4'-pentamethoxyflavone (2) by comparing its chemical (mp) and spectral (IR, NMR) data with that reported in the lit. [4].

Oxidation of candidol dimethyl ether (50 mg) in KOH (100 mg) and EtOH (50 ml) at 100° and extraction with NaHCO₃ gave a compound (10 mg), mp 182–184°, identified as anisic acid. The residue crystallized from alcohol as pale yellow prisms (15 mg), mp 76–77° (lit. [5] mp 77–78°), identified as 2-hydroxy-4,5,6, ω -tetramethoxyacetophenone by direct comparison (mmp, co-TLC and co-IR) with a synthetic sample.

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